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BA/CA RATIOS IN OTOLITHS OF SOUTHERN BLUEFIN TUNA (THUNNUS MACCOYII) AS A BIOLOGICAL TRACER OF UPWELLING IN THE GREAT AUSTRALIAN BIGHT

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Key words: southern bluefin tuna, otolith elemental composition, migratory environmental history, upwelling.

ABSTRACT

The southern bluefin tuna (*Thunnus maccoyii*) is a longlived, large, and highly migratory marine fish in the Indian Ocean. They can live up to 40 years and migrate to the Great Australian Bight in the summer when 1-4 years old. The Great Australian Bight is characterized as the most productive coastal upwelling zone in southern Australia and is the largest area of cool-water carbonate sedimentation in the world. The barium (Ba) level is poor in the open ocean but rich in the upwelling area. This study used otolith Ba/Ca ratios as a natural tag to confirm that southern bluefin tuna seasonally occupy the upwelling area. Southern bluefin tuna were collected from the central Indian Ocean and the spawning ground between the island of Java, Indonesia and northwestern Australia. The temporal variation of trace elements in otoliths of the specimen was measured by laser ablation inductively coupled plasma mass spectrometry. Otolith Ba/Ca ratios were significantly elevated in the summer growth zone, which corresponds to the upwelling season when juvenile tuna enter the Great Australian Bight at the age of 1-4 years old. Although almost all of the mature southern bluefin tuna collected in the spawning ground had previously migrated to the Great Australian Bight upwelling area as juveniles, some fish collected from the Central Indian Ocean didn't migrate to the Great Australian Bight upwelling area, perhaps because they are a vagrant population and may contribute less to the spawning stock.

I. INTRODUCTION

The southern bluefin tuna (SBT, *Thunnus maccoyii*) is a long-lived, large, and highly migratory marine fish. They can live for up to 40 years and grow up to 2 m in length and over 200 kg in weight [18]. The tuna is mainly distributed between 30°-50°S latitude throughout three Oceans in the southern hemisphere [8]. Its only spawning ground is located around 7°-20°S latitude and 100-125°E longitude between the island of Java, Indonesia and northwestern Australia [8]. The SBT spawns during the austral summer between September and March [43]. After hatching, the post-larval fishes disperse southwards along the western coast of Australia with the tropical Leeuwin Current to the inshore waters between Perth and Esperance [8]. The SBT arrives in waters of southwestern Australia during their juvenile stage, and they migrate east along the southern coast towards nursery grounds in the Great Australian Bight (GAB). The migration of juvenile SBT in the GAB was confirmed by tracking with archival tags [21]. They migrate to the east or west of Australia within the latitudes of 30-50°S during autumn, and return to the GAB in the spring before 4 years old [19]. The GAB, the largest area of cool-water carbonate sedimentation in the world, is characterized as the most productive coastal upwelling zone in southern Australia [26]. In addition, the GAB plays an important role in primary production for Australian fisheries. After 5 years old, almost all SBT have recruited to the adult stock in the open ocean. They then circumglobally disperse to temperate feeding grounds between 30°S and 50°S [8]. After maturation at 8-12 years old, the tuna only migrates between their tropical spawning ground in the tropical zone and feeding grounds in the temperate zone [20].

Otoliths function for the hearing and balance of the fish. They are mainly composed of calcium carbonate with a minor

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Table 1. Mean (± SD) and range of fork length, body weight, and age of southern bluefin tuna used for otolith trace element analysis, which were collected from the central Indian Ocean (CIO) and the spawning ground South of Java (SPAWN). n: sample size.

Sampling sites	Sampling Date	n	Fork length (cm)	Body weight (kg)	Age (yrs)
CIO	14 July - 3 August 2004	14	$122.7 (\pm 22.9)$	34.1 (± 21.4)	$10.1 (\pm 7.7)$
$(31-33°S, 71-77°E)$			$91 - 170$	$12 - 79$	4 - 27
SPAWN	1 Jan & 3 Feb 2005		$172.9 \ (\pm 8.6)$	$98.9 \ (\pm 20.6)$	$16.3(\pm 3.2)$
$(10-20°S, 110-120°E)$			160 - 189	75 - 137	$13 - 25$

Fig. 1. Sampling sites of southern bluefin tuna in the central Indian Ocean (CIO, 31-33°S, 71-77°E) and the spawning ground south of Java (SPAWN, 10-20°S, 110-120°E) [8].

organic matrix and a few trace elements. The otoliths are deposited in a daily and annual schedule that is synchronized with the photoperiod and seasonal temperature variation. As such, it yields recognized growth checks, which are daily growth increments or annuli in the otolith. This allows fish age determination at daily or annual cycles. At least 31 elements have been found to be deposited in the otolith during the growth of fish [7]. Otolith growth increments are metabolically inert once deposited and become a permanent record of fish life history. The otolith trace element such as barium (Ba) is taken up from the ambient water and is positively correlated to ambient water concentrations [3]. The barium in estuaries and bays mostly originates from freshwater but both coastal and oceanic upwelling areas are rich with barium. An increased barium concentration in fish otoliths might be a signature of oceanic upwelling [3, 9]. Thus, it should be possible to determine whether fish have encountered an upwelling area by examining the temporal change of the barium concentration in their otoliths.

The archival tag has been used to track the population structure and migration of juvenile SBT in the GAB [21] and northern bluefin tunas in the Atlantic and Pacific Oceans [4, 25]. The population structure, migratory environmental history and physiology of the SBT has also been studied by analyzing the otolith elemental composition by EPMA (electron probe microanalyzer) [35] and by laser-ablation inductively coupled plasma mass spectrometry (LA-ICPMS) [44]. However, knowledge of the migratory behavior and habitat use of the SBT in association with its environment remains fragmented. And few studies have explored the otolith elemental signature of the tuna in relation to oceanic upwelling [39].

This study tested whether the Ba in otoliths is a reliable natural tag to explore the migration of the SBT to upwelling areas. Laboratory and field studies of otolith Ba/Ca ratios in marine fish in relation to fish migration to an upwelling area have shown a positive relationship between the Ba/Ca concentration in the ambient water and otolith. Thus, we analyzed the temporal change of Ba/Ca ratios in the otolith of SBT by LA-ICPMS, and examined the life history relationship between the migration behavior of SBT and the variation in otolith microchemistry.

II. MATERIAL AND METHODS

1. Specimen Collection

The SBT was collected from two different habitats - their feeding ground in the central Indian Ocean (CIO) and their spawning ground in the waters between southern Java and northwestern Australia (SPAWN). Otolith trace elements were analyzed by ICPMS for 14 SBT from the CIO and for 15 SBT from the SPAWN (Fig. 1). The mean length, weight, and age of the specimens are shown in Table 1. Otoliths of SBT from the CIO were removed immediately after capture in the fishing boat by the observer with a battery powered hole-saw drill, while those from the SPAWN were removed in the fish market 1 to 7 days after capture. After collection, the otolith specimens were cleaned with de-ionized water, air-dried, and then stored in Eppendorf microcentrifuge tubes for age determination and otolith microchemistry analysis.

2. Otolith Preparation for Microchemistry Analyses by LA-ICPMS

For the analysis of trace elements, each otolith was cleaned with 5% H_2O_2 to remove all remaining organic material from the otolith surface, ultrasonically cleaned with de-ionized water, oven-dried overnight at 60°C, and embedded in epofix resin and transversely sectioned with a low speed saw (Isomet, Buehler) into slices approximately 300 µm thick with the primordium in the middle. The sectioned otolith was ground

Fig. 2. (upper) The LA-ICPMS measurement axis along the long arm of the otolith from the primordium to the otolith edge and (lower) the annuli in the otolith of a 17-year-old southern bluefin tuna $(FL = 172$ cm). P: primordium, $1st$ and $2nd$ IPs: first and second **inflection points, solid circles: annuli.**

with 2000 grit sandpaper followed by 2400 grit carborundum until the primordium appeared. Finally, the otolith sections were polished with 0.05 μ m alumina paste to smooth the surface.

The elements in the otoliths were measured from the primordium to the edge of the long arm (Fig. 2) by a high resolution LA-ICPMS (Finnigan MAT ELEMENT 2, Thermo Electron corp., Bremen, Germany) connected with a Merchantek LUV 266^{TM} Nd: YAG UV laser microprobe (New Wave Research, Inc). The laser was pulsed at a repetition rate of 20 Hz, at a scan speed of 15 μ m ⋅ sec⁻¹ with an ablation transect diameter of approximately 150 μ m. The elements, t^{23} Na, t^{24} Mg, t^{44} Ca, t^{55} Mn, t^{88} Sr, and t^{138} Ba, were measured because they always remain at least 10 times higher than the background levels. The setting took about 2.46 sec to produce one data point, which represented approximately 37 µm on the otolith transect. Standards (NIST 612) were collected before each series, with each series comprising 2-3 otoliths. At the start of each otolith analysis, background counts were collected for 30 seconds, and the average was then subtracted from the sample counts to correct for the background level. The ablation chamber was purged for 60 seconds after sampling each otolith. All measurement data were expressed as ratios of element to Ca concentrations (ppm \cdot ppm⁻¹) by estimating the relative response factor of the instrument to the known concentration in the standard (NIST 612).

3. Otolith Annulus and Microstructure Examination

The annulus (opaque zone) on the polished otolith sections of SBT was identified from a photograph taken under transmitted light with a compound microscope equipped with a digital camera. The SBT otolith annulus consists of a dark opaque zone and a light translucent zone under transmitted light (Fig. 2). The age of the tuna was determined by counting the number of annuli along the ventral arm, following the

manual for age determination of southern bluefin tuna *Thunnus maccoyii* [1]. The terminology describing the otolith used in this study followed Rees *et al*. [38]. Respectively, the first and second inflection points in the otolith were identified to examine the temporal change in the otolith elemental signature during the early life stage of the fish and during maturation.

4. Data Analyses

The general spatio-temporal variation in otoliths of the six elements examined (Na, Mg, Ca, Mn, Sr, and Ba) was described by Wang *et al*. [44]. The spatial variation of barium in otoliths has now been demonstrated for a variety of species [12, 23]. A positive relationship exists for barium between the otolith and ambient water, where the water temperature and salinity do not vary greatly [31]. Ambient data for barium can be used to predict the spatial and temporal variation of barium in otoliths. Likewise, barium concentrations in the otolith can potentially be used to reconstruct histories of the ambient water mass [22]. In this study, we focused on the timing of the peak Ba/Ca ratios in the otolith in relation to fish age and the relative position of the annulus to understand the timing of SBT recruitment to the upwelling area of the GAB. The criterion in the judgement of the otolith Ba/Ca ratio as a signal of upwelling for the tuna was determined from the average otolith Ba/Ca ratio (3.7×10^{-6}) of 14 marine spices reviewed by Campana [7] and that of other marine fish species (less than 4.0×10^{-6} [16, 22, 41]. Otolith Ba/Ca ratios greater than $4.0 \times$ 10⁻⁶ were identified as a signal of upwelling. The age and season when the tuna migrated to the upwelling area was determined from the position of the peak Ba/Ca ratio and the annulus of the otolith. The patterns of migration to the upwelling area were classified into 7 types according to the occurrence of peak Ba/Ca ratios, and the type frequency was also compared between CIO and SPAWN by life stage. Life stage I refers to the larval stage (0 to 55 days old), stage II refers to the juvenile stage (55 days to 1 year old), stage III refers to the juvenile to sub-adult stage (1 to 4 years old), and stage IV refers to the sub-adult to adult stage (more than 4 years old), see Table 2.

III. RESULTS

1. Seasonal Occurrence of Otolith Annuli

The first inflection point in the otoliths of SBT occurred between the primordium and first annulus and corresponds to the transition from larval to juvenile life stage (Fig. 2). Otolith annual growth increments were smaller after the second inflection point, which corresponds with sexual maturation at ages 8-12, indicating a reduction in growth rate after maturation (Fig. 2).

The marginal increment in SBT otoliths from the last annulus (opaque zone) to the otolith edge was different between CIO and SPAWN (Fig. 3). A translucent zone was present in SBT collected from the spawning ground in February of the austral summer (Fig. 3(a)), but an opaque zone appeared just **Table 2. Migratory type of southern bluefin tuna based on the occurrence of peak otolith Ba/Ca ratios at different life stages. A dot indicates the presence of a Ba: Ca peak, X indicates no peak during an otolith stage.**

	Occurrence types of upwelling signature in otolith						
		by life stage					
Type							
R			X				
		Y					
	X						
E							
	X	X					

Stage I: larval stage (0-55 days old); Stage II: larval-juvenile stage (55 days-1 year old); Stage III: juvenile stage (1-4 years old); Stage IV: juvenile-adult stage (4-10 years old).

Fig. 3. Comparison between sampling seasons of the marginal increment from the last annulus (opaque zone) to the edge of southern bluefin tuna otoliths. (a) Collected in southern Java in February 2005 (austral summer), FL = 165 cm; (b) collected from the CIO in July 2004 (austral winter), FL = 151cm. Solid circles: annuli (opaque zones). (a) and (b) are magnified from (c) and (d).

at the otolith edge for SBT collected from the CIO in July of the austral winter (Fig. 3(b)). This indicated that an opaque zone was deposited in the austral winter and a translucent zone in the summer.

2. Migratory Types of SBT and Occurrence Season of Upwelling Signal in Otolith

The migratory patterns of SBT in the Great Australian Bight upwelling area as indicated by the upwelling signal of otolith peak Ba/Ca ratios were divided into 7 types (Table 2). Type A: peak Ba/Ca ratios appeared in all life stages I, II, III and IV (Figs. 4(a) and (h)), indicating that the fish encountered barium-rich inshore, coastal waters or upwelling during the larval and juvenile stages. The percentage of Type A was about 21.4% in the CIO and 6.7% in the SPAWN. Fish of Types B and C showed an upwelling signal in stage I but not at stages II and III because no peak Ba/Ca ratios were found between the first inflection and first annulus (Stage II) or in the juvenile stage at 1-4 years old (Stage III) (Figs. 4(b), (c), and (i)). Types B and C were about 14.2% in the CIO and 13.3% in the SPAWN. For types D, E, and F no upwelling signal occurred at stages I and II (Figs. $4(d)$, (e), (f), (i), (k), and (l)), indicating that they delayed migrating to the barium-rich areas until the juvenile stage. Type E fish didn't return to the barium-rich area again. The percentage of Type D, E, and F was about 57.2% in the CIO and 80.0% in the SPAWN, respectively. Type G fish had no upwelling signal in any life stage (Fig. $4(g)$), indicating that after hatching they didn't migrate to the barium-rich area until capture. They only occurred in the CIO (7.1%).

The peak otolith Ba/Ca ratios of all 29 SBT appeared in the translucent zone of the otolith, indicating that they encountered a barium-rich area in the summer (Fig. 4).

3. Comparison of Migratory Patterns between the CIO and the SPAWN by Age Group

Type D was dominant among the 7 types of migratory patterns, reaching 60.0% for the SPAWN and 28.6% for the CIO (Table 3). This implied that most SBT (90%) migrate to the upwelling area after their juvenile stage (ages 1-4). The frequency of the occurrence of peak SBT otolith Ba/Ca ratios was not consistent between CIO and SPAWN $(X^2 = 6.040,$ $p < 0.05$) (Fig. 5), indicating the migratory pattern was different for the SBT between sampling location. Although only a low percentage of larval stage (before the first inflection) tuna migrated to the upwelling area for both CIO and SPAWN (20-40%), the percentage was higher in SPAWN (60%) than in CIO (30%) for age groups 2-3 and 3-4 ($X^2 = 4.441$ and 4.209, $p < 0.05$).

IV. DISCUSSION

1. Linkage between the Peak of Otolith Ba/Ca Ratio and Upwelling Events

Juvenile SBT enter southern Australian waters and aggre-

1

 2^{3}

Fig. 4. Temporal changes in otolith Ba/Ca ratios of southern bluefin tunas collected from the central Indian Ocean (CIO, Types A-G) and from the spawning ground south of Java (SPAWN, Types A and C-F). The Types A-G are described in Table 2. The locations of annuli with numerals and of peak Ba/Ca ratios greater than 4.0 × 10⁻⁶ (the criterion as upwelling signal, red band) are indicated by black and grey arrows, respectively. Age is indicated with the sampling site of each fish. The 1st inflection is shown in Fig. 2.

Ba/Ca

CIO-6 yrs

Table 3. Comparison of the occurrence frequency of the type of upwelling signature in otolith of southern bluefin tuna between Central Indian Ocean (CIO, $n = 14$) and spawning area (SPAWN, $n = 15$). **Total sample size = 29. Types A-G are described in Table 2.**

Fig. 5. Comparison of the frequency of occurrence of peak Ba/Ca ratios in southern bluefin tuna otoliths, by age group, for the central Indian Ocean (CIO) and the spawning ground south of Java (SPAWN).

gate in the Great Australian Bight (GAB) during the austral summer, then disperse to the east or west of Australia within latitudes 30-50°S during the autumn and return to the GAB in the spring [19]. The GAB is characterized as the most productive coastal upwelling zone in southern Australia and is the largest area of cool-water carbonate sedimentation in the world [26]. The otolith Ba/Ca ratio of SBT peaked during the austral summer, mostly at the ages of 1-4 years old, which corresponds to the age and the season of SBT appearing in the GAB. This suggests that the otolith Ba/Ca ratio is a reliable natural tag for recording the migratory environmental history of SBT to the upwelling area (GAB).

2. Occurrence Season of the Peak Ba/Ca Ratios in Otolith

Studies of the gut contents suggest that SBT migrated to the GAB to forage [29, 45]. The primary food items of juvenile SBT during the summer in the GAB are pilchards (*Sardinops*

sagax) and anchovy (*Engraulis australis*). These forage fish spawn in the upwelling area during the austral summer and autumn [45], which might attract SBT to migrate to the GAB during the austral summer. This supports the idea that the peak SBT otolith Ba/Ca ratios were deposited during their foraging migration to the GAB during the summer season. The variation in the magnitude of peak otolith Ba between years might be due to the inter-annual change in the strength of the upwelling.

The otolith Ba/Ca ratios of other fish are weakly affected by temperature, which might influence the estimation of the upwelling effect on the peak otolith Ba/Ca ratios, and subsequently the migratory environmental history of the fish [3, 22, 30]. Salinity may also influence the incorporation of barium into the otolith, particularly in diadromous or freshwater fishes [11, 22]. The SBT lives in the open ocean which has a comparatively constant environment relative to that of diadromous or freshwater fishes in the estuary or freshwater. Thus, salinity might not greatly influence the otolith Ba/Ca ratios of tunas in the open ocean. Barium concentration varies positively with the environmental water chemistry [13, 30]. We have no data to verify whether the summer peak in SBT otolith Ba/Ca ratios was influenced by seasonal changes in water temperature. If the effect of the temperature on the barium uptake of the fish was significant, then another peak otolith Ba/Ca ratio would be expected when the SBT migrated between the tropical spawning area south of Java after maturation at the age of 8-12 years [20]. However, no such seasonal peak Ba/Ca ratio was found for most SBT, suggesting a negligible temperature effect on otolith Ba/Ca ratios.

3. Individual Variation in Peak Otolith Ba/Ca Ratios

The peak of SBT otolith Ba/Ca ratios occurred mostly at the age of 1-4 years old, which corresponds to the age of the SBT migration to the upwelling area of the GAB. However, the age of the occurrence of peak otolith Ba/Ca ratios varies among individuals, which might indicate that the timing at recruitment to the upwelling area of GAB differs among individuals. Fish of Types A-C had peak otolith Ba/Ca ratios before the first inflection at approximately 45–55 days old which corresponds to the life-stage transition from larva to juvenile and a habitat shift away from the inshore nursery area in northwestern Australia. Two types of upwelling occur in the Indian Ocean, wind-driven coastal upwelling and current-induced oceanic upwelling. Both types of upwelling may have a different influence on *T. maccoyii*. Larvae < 45–55 days old (or before the first inflection point) are usually distributed between southern Java and north-western Australian but otolith Ba:Ca concentration ratios were low at this age, suggesting that no upwelling happened in this location as found by Wyrtki (1962) [47]. This indicated that SBT might encounter Ba-rich waters in the spawning area at the larval-juvenile stage transition. The only known SBT spawning ground is located between southern Java and northwestern Australia (about 7°-20°S and 100-125°E) where several currents occur, including the Indonesian through flow (ITF), South Java current (SJC), South Equatorial current (SEC), Eastern Gyral current (EGC) and the Leeuwin current (LC). The mixing of these currents might produce an upwelling of the Ba-rich deep water to the surface and enable a high Ba uptake by some larval SBT. There is an upwelling along Java and Sumatra near the spawning area [5, 36]. However, the appearance of the upwelling signature was controlled by the annual monsoon and remotely by ENSO (El Niño-Southern Oscillation) [42]. This can explain the elevated Ba/Ca ratios in the otolith of SBT at the juvenile stage. The larval SBT is usually transported from the spawning ground by the southward-flowing Leeuwin Current to the Great Australian Bight along the west Australian coast where no upwelling occurs [47]. The productivity in the coastal waters of western Australian is low because of downwelling and the polewardflowing Leeuwin Current [14, 24]. The southward Leeuwin Current is characterized by warm and low salinity waters that seasonally change with maximum flows in autumn and winter [34]. Sardine eggs and larvae are abundant along the southwestern coasts of Australia from June to August and from December to February, which is linked to the coastal upwelling [15, 17, 33]. The occurrence of some SBT with peak otolith Ba/Ca ratios before age-1 might be the result of coastal upwelling along the southwestern Australia coast. A higher barium upwelling signature helped distinguish the different nursery grounds of the northern bluefin tuna (*Thunnus orientalis*) [39].

In addition, stable isotopes of oxygen in the otolith have been validated for reconstructing the temperature histories of fish [40]. The $\delta^{18}O$ values in otoliths were negatively correlated with the ambient temperatures experienced by the fish. After spawning the larval SBT migrated to the southern temperate zone from the tropics. Shiao *et al*. (2009) used the composition of both O and C isotopes to reconstruct the migratory temperature history of SBT and show a correlation between otolith $\delta^{18}O$ values and the ambient temperature for SBT less than one-year-old [40]. The otolith $\delta^{18}O$ values increased gradually as the temperature decreased when fish migrated from the spawning ground to the nursery ground. After age one, the thermal conservation ability of the fish developed gradually, however, the seasonal change of $\delta^{18}O$ values in otolith was still obvious (see Fig. 4 in Shiao *et al*. 2009) [40]. The $\delta^{18}O$ values between two annual otolith increments in the summer zone were higher than those in the winter zone at ages less than 4 years. This indicated the fish migrated to a low temperature upwelling area during the high temperature summer season. The GAB is the largest cool-water carbonate upwelling area in the world. The match between the temperature history of the SBT as revealed from both otolith oxygen isotopes and Ba/Ca ratios indicates that the otolith Ba/Ca of SBT can be used to examine fish habitat residency in areas of oceanic upwelling.

The Ba concentration in the surface waters of the Southern Ocean ranges between 4.2-5.5 μ g ⋅ L⁻¹, which is similar to that in the GAB upwelling area $(5-6 \mu g \cdot L^{-1})$ [22, 27]. Thus, the peak Ba/Ca ratios in otolith of SBT older than 5 years might be due to fish migrating to the southern ocean or polar front. Between age 5 and maturation at the age of 8–12 years, SBT migrate from the Great Australian Bight eastward and westward to the circumglobal feeding grounds between 30 and 50°S in the open ocean. The area 30–50°S is located at the boundary between subarctic and central Indian Ocean waters, which is a divergence zone where Ba-rich deep water may upwell to the surface layer of the ocean. Thus, a peak otolith Ba:Ca ratio may occur annually, but not regularly, in *T. maccoyii* after maturation. The difference in amplitude of the Ba:Ca ratios might indicate annual changes in the strength of the upwelling. The SBT from the CIO where no peak otolith Ba/Ca occurred between the ages of 1-4 years might indicate that some larvae entered the central Indian Ocean directly via the South Equatorial current without entering the GAB after hatching. The higher percentage of SBT with peak otolith Ba/Ca ratios from the SPAWN than from the CIO might indicate that inhabiting areas of upwelling during the juvenile stage might contribute to improved spawning stock recruitment.

V. CONCLUSION

We conclude that otolith Ba/Ca ratios and associated age determinations can be used as a biological tracer to identify the age and season of SBT migrating to an upwelling area. Seasonal movements in the GAB might be linked to food availability, which is influenced by the upwelling. The higher percentage of SBT on the spawning ground with peak otolith Ba/Ca ratios indicates that inhabiting areas of upwelling may improve spawning stock recruitment.

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REFERENCES

- 1. Anonymous, "A manual for age determination of southern bluefin tuna *Thunnus maccoyii*: otolith sampling, preparation and interpretation," Report of the Direct Age Estimation Workshop, Victoria, Australia (2002).
- 2. Ashford, J. R., Arkhipkin, A. I., and Jones, C. M., "Otolith chemistry reflects frontal systems in the Antarctic Circumpolar Current," *Marine Ecology Progress Series*, Vol. 351, pp. 249-260 (2007).
- 3. Bath, G. E., Thorrlod, S. R., Jones, C. M., Campana, S. E., McLaren, J. W.,

and Lam, J. W. H., "Strontium and barium uptake in aragonitic otoliths of marine fish," *Geochimica et Cosmochimica Acta*, Vol. 64, pp. 1705-1714 (2000).

- 4. Block, B. A., Teo, S. L. H., Walli, A., Boustany, A., Stokesbury, M. J. W., Farwell, C. J., Weng, K. C., Dewar, H., and Williams, T. D., "Electronic tagging and population structure of Atlantic bluefin tuna," *Nature*, Vol. 434, pp. 1121-1127 (2005).
- 5. Bray, N. A., Hautala, S., Chong, J., and Pariwono, J., "Large-scale sea level, thermocline, and wind variations in the Indonesian throughflow region," *Journal of Geophysical Research*, Vol. 101, pp. 12239-12254 (1996).
- 6. Brown, R. and Severin, K. P., "Elemental distribution within polymorphic inconnu (*Stenodus leucichthys*) otoliths is affected by crystal structure," *Canadian Journal of Fisheries and Aquatic Sciences*, Vol. 56, pp. 1898-1903 (1999).
- 7. Campana, S. E., "Chemistry and composition of fish otolith: pathways, mechanisms and applications," *Marine Ecology Progress Series*, Vol. 188, pp. 263-297 (1999).
- 8. Caton, A. E., "Review of aspects of southern bluefin tuna biology, population and fisheries. In World meeting on stock assessment of bluefin tunas: strengths and weaknesses," in: Deriso, R. B. and Bayliff, W. H. (Eds.), *Inter-American Tropical Tuna Commission, Special Report*, Vol. 7, pp. 181-350 (1991).
- 9. Clarke, A. D., Lewis, A., Telmer, K. H., and Shrimpton, J. M., "Life history and age at maturity of an anadromous smelt, the eulachon *Thaleichthys pacificus*," *Journal of Fish Biology*, Vol. 71, pp. 1479-1493 (2007).
- 10. Coffey, M., Dehairs, F., Collette, O., Luther, G., Church, T., and Jickells, T., "The behaviour of dissolved barium in estuaries," *Estuarine, Coastal and Shelf Science*, Vol. 45, pp. 113-121 (1997).
- 11. De Vries, M. C., Gillanders, B. M., and Elsdon, T. S., "Facilitation of barium uptake into fish otoliths: influence of strontium concentration and salinity," *Geochimica et Cosmochimica Acta*, Vol. 69, pp. 4061-4072 (2005).
- 12. Elsdon, T. S. and Gillanders, B. M., "Reconstructing migratory patterns of fish based on environmental influences on otolith chemistry," *Reviews in Fish Biology and Fisheries*, Vol. 13, pp. 219-235 (2003).
- 13. Elsdon, T. S. and Gillanders, B. M., "Consistency of patterns between laboratory experiments and field collected fish in otolith chemistry: an example and applications for salinity reconstructions," *Marine and Freshwater Research*, Vol. 56, pp. 609-617 (2005).
- 14. Feng, M., Meyers, G., Pearce, A., and Wijffels, S., "Annual and interannual variations of the Leeuwin Current at 32°S," *Journal of Geophysical Research*, Vol. 108, pp. 19-1-19-21 (2003).
- 15. Fletcher, W. J., Tregonning, R. J. and Sant, G. J., "Interseasonal variation in the transport of pilchard eggs and larvae off southern Western Australia," *Marine Ecology Progress Series*, Vol. 111, pp. 209-224 (1994).
- 16. Fowler, A. J., Gillanders, B. M., and Hall, K. C., "Relationship between elemental concentration and age from otoliths of adult snapper (*Pagrus auratus*, Sparidae): implicationsfor movement and stock structure," *Marine and Freshwater Research*, Vol. 56, pp. 661-676 (2005).
- 17. Gaughan, D. J., Neira, F. J., Beckley, L. E., and Potter, I. C., "Composition, seasonality and distribution of the ichthyoplankton in the lower Swan Estuary, south-western Australia," *Australian Journal of Marine and Freshwater Research*, Vol. 41, pp. 529-543 (1990).
- 18. Gunn, J. S., Clear, N. P., Carter, T. I., Rees, A. J., Stanley, C. A., Kalish, J. M., and Johnston, J. M., "Age and growth of southern bluefin tuna 1995 Report on Research," Commission for the Conservation of Southern Bluefin Tuna CCSBT Scientific Meeting, CCSBT, Schimuzn, Japan (Pap. No. 95/8) (1995).
- 19. Gunn, J. S. and Young, J., "Environmental determinants of the movement and migration of juvenile southern bluefin tuna," in: *Fish Movement and Migration Workshop Proceeding*, Australian Society for Fish Biology. Sydney (1996).
- 20. Gunn, J. S., Farley, J. H., and Clear, N. P., "The age distribution and relative strength of cohorts of SBT on the spawning grounds," Commis-

sion for the Conservation of Southern Bluefin Tuna, CCSBT/9807/39 (1998).

- 21. Gunn, J. and Block, B., "Advances in acoustic, archival, and satellite tagging of tunas," *Fish Physiology*, Vol. 19, pp. 167-224 (2001).
- 22. Hamer, P. A., Jenkins, G. P., and Coutin, P., "Barium variation in *Pagrus auratus* (Sparidae) otoliths: A potential indicator of migration between an embayment and ocean waters in south-eastern Australia," *Estuarine, Coastal and Shelf Science*, Vol. 68, pp. 686-702 (2006).
- 23. Hamer, P. A. and Jenkins, G. P., "Comparison of spatial variation in otolith chemistry of two fish species and relationships with water chemistry and otolith growth," *Journal of Fish Biology*, Vol. 71, pp. 1035-1055 (2007).
- 24. Hanson, C. E., Pattiaratchi, C. B., and Waite, A. M., "Sporadic upwelling on a downwelling coast: Phytoplankton responses to spatially variable nutrient dynamics off the Gascoyne region of Western Australia," *Continental Shelf Research*, Vol. 25, pp. 1561-1582 (2005).
- 25. Itoh, T., Tsuji, S., and Nitta, A., "Migration patterns of young Pacific bluefin tuna (*Thunnus orientalis*) determined with archival tags," *Fishery Bulletin*, Vol. 101, pp. 514-534 (2003).
- 26. James, N. P., Bone, Y., Collins, L. B., and Kyser, T. K., "Surficial sediments of the Great Australian Bight: Facies dynamics and oceanography on a vast cool-water carbonate shelf," *Journal of Sedimentary Research*, Vol. 71, pp. 549-567 (2001).
- 27. Jeandel, C., Dupre, B., Lebaron, G., Monnin, C., and Minister, J. F., "Longitudinal distributions of dissolved barium, silica and alkalinity in the western and southern Indian Ocean," *Deep-Sea Research Part II: Topical Studies in Oceanography*, Vol. 43, pp. 1-31 (1996).
- 28. Kalish, J. M., "Otolith microchemistry: validation of the effects of physiology, age and environment on otolith composition," *Journal of Experimental Marine Biology and Ecology*, Vol. 132, pp. 151-178 (1989).
- 29. Kemps, H., Totterdell, J., Gill, H., and Nishida, T., "Preliminary analysis on the diet and feeding ecology of juvenile southern bluefin tuna, *Thunnus maccoyii*, in relation to the southern coastal waters of Western Australia," in: *Report to the Tenth Workshop, SBT Recruitment and Monitoring Program*, CSIRO Marine Research, Hobart (1998).
- 30. Martin, G. B. and Thorrold, S. R., "Temperature and salinity effects on magnesium, manganese, and barium incorporation in otoliths of larval and early juvenile spot *Leiostomus xanthurus*," *Marine Ecology Progress Series*, Vol. 293, pp. 223-232 (2005).
- 31. Martin, G. B. and Wuenschel, M. J., "Effect of temperature and salinity on otolith element incorporation in juvenile gray snapper *Lutjanus griseus*," *Marine Ecology Progress Series*, Vol. 324, pp. 229-239 (2006).
- 32. Melancon, S., Fryer, B. J., Ludsin, S. A., Gagnon, J. E., and Yang, E., "Effects of crystal structure on the uptake of metals by lake trout (*Salvelinus namaycush*) otoliths," *Canadian Journal of Fisheries and Aquatic Sciences*, Vol. 62, pp. 2609-2619 (2005).
- 33. Muhling, B. A., Beckley, L. E., Gaughan, D. J., Jones, C. M., Miskiewicz, A. G., and Hesp, S. A., "Spawning, larval abundance and growth rate of *Sardinops sagax* off southwestern Australia: influence of an anomalous eastern boundary current," *Marine Ecology Progress Series*, Vol. 364, pp. 157-167 (2008).
- 34. Pearcea, A. F., Lynchb, M. J., and Hansona, C. E., "The Hillarys Transect (1): Seasonal and cross-shelf variability of physical and chemical water properties off Perth, Western Australia, 1996-98," *Continental Shelf Research*, Vol. 26, pp. 1689-1729 (2006).
- 35. Proctor, C. H., Thresher, R. E., Gunn, J. S. Mills, D. J., Harrowfield, I. R., and Sie, S. H., "Stock structure of the southern bluefin tuna *Thunnus maccoyii*: An investigation based on probe microanalysis of otolith composition," *Marine Biology*, Vol. 122, pp. 511-526 (1995).
- 36. Potemra, J. T. and Lukas, R., "Seasonal to interannual modes of sea level variability in the western Pacific and eastern Indian Oceans," *Geophysical Research Letters*, Vol. 26, pp. 365-368 (1999).
- 37. Radtke, R. L. and Shafer, D. J., "Environmental sensitivity of fish otolith microchemistry," *Australian Journal of Marine and Freshwater Research*, Vol. 43, pp. 935- 951 (1992).
- 38. Rees, A. J., Gunn, J. S., and Clear, N. P., "Age determination of juvenile southern bluefin tuna, *Thunnus maccoyii*, based on scanning electron

microscopy of otolith microincrements," CCSBT/SC/96/8 (Appendix), Second Meeting of the Commission for the Conservation of Southern Bluefin Tuna (CCSBT), CSIRO Marine Laboratories, Hobart, Tasmania (1996).

- 39. Rooker, J. R., Secor, D. H., Zdanowicz, V. S., Metrio, G. D., and Relini, L. O., "Identification of Atlantic bluefin tuna (*Thunnus thynnus*) stocks from putative nurseries using otolith chemistry," *Fisheries Oceanography*, Vol. 12, pp. 75-84 (2003).
- 40. Shiao, J. C., Yui, T. F., Høie, H., Ninnemann, U., and Chang, S. K., "Otolith O and C stable isotope compositions of southern bluefin tuna *Thunnus maccoyii* (Pisces: Scombridae) as possible environmental and physiological indicators," *Zoological Studies*, Vol. 48, pp. 71-82 (2009).
- 41. Stransky, C., Garbe-Schönberg, C.-D., and Günther, D., "Geographic variation and juvenile migration in Atlantic redfish inferred from otolith microchemistry," *Marine and Freshwater Research*, Vol. 56, pp. 677-691 (2005).
- 42. Susanto, D. W., Gordon, A. L., and Zheng, Q., "Upwelling along the coasts of Java and Sumatra and its relation to ENSO," *Geophysical Research Letters*, Vol. 28, pp. 1599-1602 (2001).
- 43. Ueyanagi, S., "The spawning of the southern bluefin tuna (*Thunnus maccoyii*) as indicated by the occurrence of its larvae," *Bulletin Far Seas Fisheries Research Laboratory*, Vol. 1, pp. 1-4 (1969).
- 44. Wang, C. H., Lin, Y. T., Shiao, J. C., You, C. F., and Tzeng, W. N., "Spatio-temporal variation in the elemental compositions of otoliths of southern bluefin tuna *Thunnus maccoyii* in the Indian Ocean with reference to ontogenetic effect and environmental history," *Journal of Fish Biology*, Vol. 75, pp. 1173-1193 (2009).
- 45. Ward, T. M., McLeay, L. J., Dimmlich, W. F., Rogers, P. J., McClatchie, S., Matthews, R., Kampf, J., and Van Ruth, P. D., "Pelagic ecology of a northern boundary current system: effects of upwelling on the production and distribution of sardine (*Sardinops sagax*), anchovy (*Engraulis australis*) and southern bluefin tuna (*Thunnus maccoyii*) in the Great Australian Bight," *Fisheries Oceanography*, Vol. 15, pp. 191-207 (2006).
- 46. Wolgemuth, K. and Broecker, W., "Barium in sea water," *Earth and Planetary Science Letters*, Vol. 8, pp. 372-378 (1970).
- 47. Wyrtki, K., "The upwelling in the region between Java and Australia during the south-east monsoon," *Australian Journal of Marine and Freshwater Research*, Vol. 13, pp. 217-225 (1962).